Preparation of Nuclear Matrix on Slide for Immunofluorescence

- affix cells onto poly-L-lysine-coated coverslips as per standard protocol (e.g., cytospin)
- wash cells with PBS, 5 min
- incubate cells for 5 min with PBS containing 0.5% Triton X-100 (TX-100)
- remove the TX-100
- add ice-cold CSK buffer containing 0.5% TX-100 and incubate 5 min
- remove the CSK/T-100
- wash briefly twice with CSK buffer
- add DNAse solution (1 mg/ml DNAse I made in CSK buffer)
- incubate for 30 min
- remove DNAse solution
- rinse and wash 2 x 5 min with PBS
- fix samples with 3% paraformaldehyde, 10 min
- continue with the regular immunofluorescence protocol

Solutions

1) CSK buffer: see protocols for buffers; add all protease inhibitors
2) DNAse I solution: make up DNAse I to 1 mg/ml in CSK buffer (will need ~250 µl / coverslip)
3) all immunofluorescence solutions